



Do galectins serve as soluble ligands for immune checkpoint receptors?

Nicolas I Torres ^{1,2} Federico G Baudou,^{1,3} Marco A Scheidegger,¹ Tomás Dalotto-Moreno,¹ Gabriel A Rabinovich ^{1,4}

To cite: Torres NI, Baudou FG, Scheidegger MA, *et al.* Do galectins serve as soluble ligands for immune checkpoint receptors? *Journal for ImmunoTherapy of Cancer* 2024;**12**:e008984. doi:10.1136/jitc-2024-008984

Accepted 27 March 2024



© Author(s) (or their employer(s)) 2024. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

¹Laboratorio de Glicomedicina, Programa de Glicociencias, Instituto de Biología y Medicina Experimental, Consejo Nacional de Investigaciones Científicas y Técnicas, Ciudad de Buenos Aires, Argentina

²Instituto de Tecnología, Universidad Argentina de la Empresa, Ciudad de Buenos Aires, Argentina

³Departamento de Ciencias Básicas, Universidad Nacional de Luján, Luján, Provincia de Buenos Aires, Argentina

⁴Departamento de Química Biológica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Ciudad de Buenos Aires, Argentina

Correspondence to

Dr Gabriel A Rabinovich;
gabyrabi@gmail.com

ABSTRACT

Hardly any set of molecules have been studied more extensively in immuno-oncology than immune checkpoint receptors and their ligands. However, with intense research both in experimental models and clinical settings, additional immune evasion mechanisms and immunosuppressive pathways are continuously being discovered. These include galectins, a family of soluble glycan-binding proteins that shape the nature and magnitude of antitumor immune responses by reprogramming the fate and function of lymphoid and myeloid cells. Extracellularly, they interact with a variety of glycosylated receptors, modulating their cell surface retention, endocytosis, segregation, and signaling. Remarkably, a growing body of evidence suggests that galectins can bind to immune checkpoint receptors, including the T cell immunoglobulin domain and mucin domain-3 (TIM-3), programmed death-1 (PD-1), lymphocyte activating gene-3 (LAG-3), cytotoxic T lymphocyte antigen-4 (CTLA-4) and the leukocyte Ig-like receptor subfamily B3 (LILRB3) and B4 (LILRB4), favoring their signaling activity and inhibitory function. However, more work is needed to dissect the precise mechanisms underlying these effects, particularly the interplay between galectins and canonical immune checkpoint ligands as well as the biochemical nature and glycan-binding dependence of these interactions. A better understanding of 'galectin-immune checkpoint' hubs will contribute to design novel immunotherapeutic modalities aimed at targeting galectin-driven circuits in a wide range of cancer settings.

INTRODUCTION

Galectins are a family of soluble glycan-binding proteins containing one or two highly conserved carbohydrate recognition domains (CRDs) that preferentially bind to lactosamine (LacNAc; Gal β 1-4NAcGlc) structures present on different cell surface receptors and the extracellular matrix.¹ Unlike other lectins such as C-type lectin receptors and sialic acid-binding immunoglobulin-type lectins (Siglecs), which are mostly transmembrane proteins,² galectins are synthesized as cytosolic proteins, reside in the cytosol or nucleus for much of their lifetime, and are released into the extracellular medium through non-classical secretory mechanisms bypassing the endoplasmic reticulum

(ER)-Golgi system. This externalization pathway may include accumulation inside extracellular vesicles and activation of inflammatory cell death pathways, including pyroptosis and necroptosis.³⁻⁶

Galectins have been classified into three groups according to their structure: (a) the 'prototype' galectins (galectin-1, galectin-2, galectin-7 to galectin-10, galectin-13 and galectin-14), which contain a single CRD that may associate to form homodimers; (b) the 'tandem-repeat' galectins (galectin-4, galectin-8, galectin-9 and galectin-12), in which two CRDs occur within a single polypeptide chain connected by a short linker peptide and (c) the 'chimera-type' galectin-3, containing a single CRD and a large amino-terminal domain that contributes to oligomerization. In the past years, galectins have been proposed to play central roles in different hallmarks of cancer.⁷ Particularly, they have emerged as key regulators of anti-tumor immune responses and novel targets of immunotherapy by shaping the fate and function of both lymphoid and myeloid cells.^{8,9} Interestingly, expression of different galectins is altered on different cell types of the tumor microenvironment, particularly, tumor cells and their associated endothelium, stromal fibroblasts, and immune cells.³ At the molecular level, they bind to a preferential set of glycosylated receptors, including the T cell receptor (TCR), the B cell receptor (BCR), integrins, cytokine receptors, tyrosine phosphatase receptors (eg, CD45) and tyrosine kinase receptors (eg, VEGFR2), forming multivalent lectin-glycan complexes, often termed lattices, that control receptor segregation, endocytosis, signaling and activation.¹⁰ Moreover, emerging evidence suggests a possible role of galectins as functional ligands of different immune checkpoint receptors present on lymphoid and myeloid cells (figure 1).

To illustrate this concept, galectin-9 has been demonstrated to play a key role in immune

Protected by copyright. including for uses related to text and data mining, AI training, and similar technologies.

J Immunother Cancer: first published as 10.1136/jitc-2024-008984 on 9 April 2024. Downloaded from <http://jitc.bmj.com/> on May 20, 2025 at Department GEZ-LTA ErasmusHogeschool.

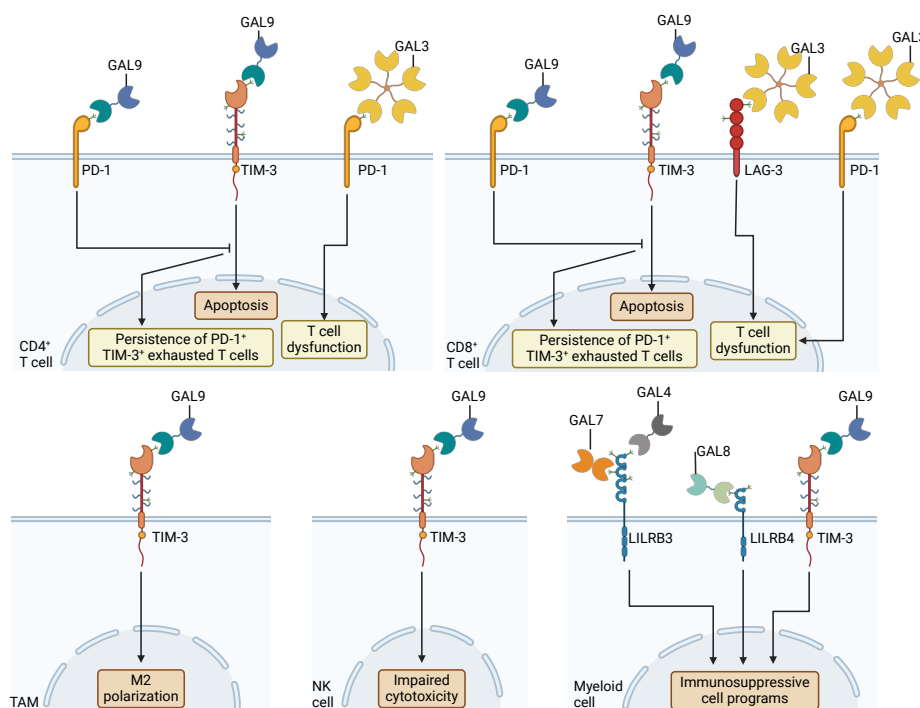


Figure 1 Galectins can bind to immune checkpoint receptors and trigger inhibitory pathways. Galectins can reprogram antitumor immune responses by shaping both lymphoid and myeloid compartments. Among the different inhibitory pathways triggered by these β -galactoside-binding proteins, a growing body of experimental evidence suggests that they can bind to a set of distinct immune checkpoint receptors including TIM-3, LAG-3, PD-1, LILRB3 and LILRB4, thus amplifying immune inhibitory circuits and facilitating tumor-immune escape in different cancer types. GAL: galectin; TAM: tumor-associated macrophages. Created with BioRender.com.

escape through mechanisms that are, at least in part, dependent on T cell immunoglobulin domain and mucin domain-3 (TIM-3) engagement.¹¹ Seminal work demonstrated that galectin-9 blunts T helper (Th)1 responses through binding to TIM-3.¹² Since then, the relevance of the TIM-3/galectin-9 axis in immuno-oncology has been studied in several tumor types, including solid tumors and hematological malignancies.^{13–19} This pathway is implicated in critical inhibitory mechanisms that limit the magnitude of anticancer responses including induction of T-cell apoptosis, promotion of myeloid-derived suppressor cells (MDSCs) and inhibition of NK cell activity.¹¹ Supporting this notion, galectin-9 impaired NK cell-mediated cytotoxicity through association with TIM-3 in acute myeloid leukemia²⁰ and contributed to the expansion of CD11b⁺Ly-6G⁺MDSCs, thus favoring immune escape.¹⁹ Interestingly, the human leukocyte antigen B (HLA-B)-associated transcript 3 (BAT-3) has been shown to counteract the TIM-3/galectin-9 axis by repressing the function of TIM-3 and protecting Th1 cells from galectin-9-mediated cell death.²¹ However, evidence has also been raised highlighting the importance of TIM-3-independent immunoregulatory programs mediated by galectin-9.²² In this regard, more recent evidence demonstrated that galectin-9 can also interact with the programmed death-1 (PD-1) pathway through N-glycan-dependent mechanisms, forming lattices that inhibit

TIM-3-mediated apoptosis and promote persistence of PD-1⁺TIM-3⁺ T cells.^{16,19} Notably, galectin-9/PD-1 interaction could not be interrupted by clinically available PD-1/PD-L1-blocking antibodies, suggesting different binding sites for this lectin on the PD-1 receptor.¹⁶ Moreover, soluble V-domain Ig-containing suppressor of T cell activation (VISTA) synthesized by acute myeloid leukemia cells has been also shown to bind galectin-9 and enhance its immunosuppressive activity by creating a multiprotein barrier which prevents granzyme B release from cytotoxic T cells.¹⁹

However, galectin-9 may also bind to the T-cell costimulatory receptor 4-1BB (CD137), a member of the tumor necrosis factor receptor superfamily, in a site distinct from its natural ligand 4-1BBL, facilitating 4-1BB aggregation, signaling, and functional activity.²³ Interestingly, this tandem-repeat lectin may also promote immunosuppression in pancreatic adenocarcinoma through binding to Dectin-1, a C-type lectin receptor commonly upregulated in tumor-associated macrophages, highlighting the ability of this lectin to modulate both innate and adaptive anti-tumor responses.²⁴ In this regard, activation of the galectin-9/TIM-3 axis can drive M2 macrophage polarization leading to poor outcome in glioma patients.¹⁴ Computational modeling and structural studies revealed the selective contribution of N- and C-terminal CRDs of galectin-9 to modulation of innate and adaptive immune functions.²⁵

In spite of its paradoxical proinflammatory and anti-inflammatory activities in different pathophysiologic settings, galectin-3 has been shown to thwart antitumor responses in different cancer settings by inducing T-cell anergy, promoting T-cell apoptosis and favoring T cell exclusion through interruption of IFN- γ -driven chemokine gradients required for T cell infiltration.^{26–28} In a model of triple negative breast cancer, tumor-secreted galectin-3 and a galectin-3-binding-protein have been shown to form a complex which interacts with CD45 receptor on T cells and induce expansion of regulatory T cells.²⁹ Interestingly, in a model of pancreatic ductal adenocarcinoma, galectin-3 has been shown to promote CD8⁺ T-cell dysfunction through binding to the lymphocyte-activation gene 3 (LAG-3).³⁰ Accordingly, blockade of the galectin-3/LAG-3 axis has been validated in T cells from multiple myeloma patients.³¹ However, the association of galectin-3 with other immune checkpoint pathways has not been studied in such detail. Wu *et al* reported increased levels of circulating anti-galectin-3 antibodies which correlated with superior clinical outcomes in patients with advanced melanoma receiving a combination of anti-CTLA-4 (ipilimumab) and anti-VEGF (bevacizumab) mAb,³² suggesting a possible cross-talk between galectin-3 and the CTLA-4 pathway. In this regard, it has been demonstrated that CTLA-4 endocytosis and inhibitory signaling are highly dependent on elongation of complex branched N-glycans and the formation of galectin-N-glycan lattices.³³ Interestingly, ongoing clinical trials are conducted using galectin-3 inhibitors to improve anti-PD-1 therapy in metastatic melanoma and head and neck squamous cell carcinoma.^{9, 34} In this regard, the functional relevance of PD-1 glycosylation and a direct interaction between PD-1 and galectin-3 have recently been demonstrated.^{35, 36} These results highlight the importance of galectin-3 as an emerging target of cancer immunotherapy either alone or in different combination modalities.^{34–37}

Like galectin-3 and galectin-9, galectin-1 has been shown to shape antitumor immune responses and foster immunosuppressive networks through modulation of lymphoid and myeloid cells.³ These effects include induction of apoptosis of Th1, Th17 and CD8⁺ T cells, expansion of CD4⁺ and CD8⁺ regulatory T cells and promotion of tolerogenic dendritic cells and immunosuppressive tumor-associated macrophages.^{38–44} Several glycosylated receptors have been shown to mediate the functional activity of this lectin, including CD45, CD7, CD43, CD146, and VEGFR2.^{1, 26, 39, 40, 42, 45} Although the association of galectin-1 with immune checkpoint pathways remains elusive, this lectin has been shown to contribute to T-cell exclusion and resistance to PD-1 blockade.^{46, 47} Interestingly, similar to galectin-3, a strong antibody response to galectin-1 has also been observed in patients with metastatic melanoma responding to anti-CTLA-4 plus anti-VEGF therapy,⁴⁸ suggesting a possible interplay between galectin-1 and these pathways. In fact, we found that galectin-1 delineates resistance to

anti-VEGF therapies, both in animal models and clinical settings.^{42, 49}

Finally, recent findings identified the leukocyte immunoglobulin-like receptor subfamily B3 (LILRB3) and the leukocyte immunoglobulin-like receptor subfamily B4 (LILRB4) as novel immune checkpoint receptors containing tyrosine-based inhibitory motifs (ITIMs) and preferentially expressed on myeloid cells.^{50, 51} Interestingly, galectin-4 and galectin-7 were discovered as high-affinity functional ligands of LILRB3, whereas galectin-8 has been demonstrated to bind LILRB4, triggering signaling of these receptors and regulating immunosuppressive myeloid cell programs.^{52, 53}

In conclusion, given the broad immunoregulatory activities of galectins in secondary lymphoid organs and tumor microenvironments, it is becoming increasingly apparent that immune checkpoint receptors could mediate at least part of their functions (figure 1). However, in spite of considerable progress, it still needs to be explored whether these proteins operate independently of canonical immune checkpoint ligands and whether they act in a coordinated fashion to control the dynamics and signaling of these receptors. Meanwhile, several experimental and clinical studies are underway to evaluate the effects of galectin-targeted therapies using small molecule glycan inhibitors, natural polysaccharides, peptidomimetics, and neutralizing monoclonal antibodies, as standalone monotherapy or in combination with other anticancer treatments, including immune checkpoint blockers.⁹

Contributors All authors contributed data for the article, made substantial contributions to discussions of content, wrote the article, and reviewed and edited the article before its submission.

Funding Work in GAR's lab is supported by grants from Ministerio de Ciencia, Tecnología e Innovación (Redes Federales de Alto Impacto, 0005) and Agencia Nacional de Promoción Científica y Tecnológica, Fondo para la Investigación Científica y Tecnológica (PICT-2017-0494 and PICT-2020-01552). The authors are thankful for generous support from Sales, Bunge & Born, Baron, Williams and Richard Lounsbury Foundations, as well as generous donations from Ferioli-Ostry and Caraballo families.

Competing interests GAR is cofounder of GALTEC S.A.S.

Patient consent for publication Not applicable.

Provenance and peer review Commissioned; externally peer reviewed.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See <http://creativecommons.org/licenses/by-nc/4.0/>.

ORCID iDs

Nicolas I Torres <http://orcid.org/0000-0001-6501-185X>

Gabriel A Rabinovich <http://orcid.org/0000-0002-0947-8735>

REFERENCES

- Troncoso MF, Elola MT, Blidner AG, *et al*. The universe of Galectin-binding partners and their functions in health and disease. *J Biol Chem* 2023;299:105400.
- Smith BAH, Bertozzi CR. The clinical impact of Glycobiology: targeting selectins, Siglecs and mammalian Glycans. *Nat Rev Drug Discov* 2021;20:217–43.

- 3 Rabinovich GA, Conejo-García JR. Shaping the immune landscape in cancer by Galectin-driven regulatory pathways. *J Mol Biol* 2016;428:3266–81.
- 4 Thijsen VL, Rabinovich GA, Griffioen AW. Vascular Galectins: regulators of tumor progression and targets for cancer therapy. *Cytokine Growth Factor Rev* 2013;24:547–58.
- 5 Russo AJ, Vasudevan SO, Méndez-Huergo SP, et al. Intracellular immune sensing promotes inflammation via Gasdermin D-driven release of a lectin Alarmin. *Nat Immunol* 2021;22:154–65.
- 6 Johannes L, Jacob R, Leffler H. Galectins at a glance. *J Cell Sci* 2018;131:jcs208884.
- 7 Girotti MR, Salatino M, Dalotto-Moreno T, et al. Sweetening the hallmarks of cancer: galectins as multifunctional mediators of tumor progression. *J Exp Med* 2020;217:e20182041.
- 8 Navarro P, Martínez-Bosch N, Blidner AG, et al. Impact of Galectins in resistance to anticancer therapies. *Clin Cancer Res* 2020;26:6086–101.
- 9 Mariño KV, Cagnoni AJ, Croci DO, et al. Targeting Galectin-driven regulatory circuits in cancer and fibrosis. *Nat Rev Drug Discov* 2023;22:295–316.
- 10 Mkhikian H, Sy M, Dennis JW, et al. The Galectin-lattice: a Decoder of bio-equivalent Glycans. *Glycoforum* 2021;24.
- 11 Sakuishi K, Jayaraman P, Behar SM, et al. Emerging Tim-3 functions in antimicrobial and tumor immunity. *Trends Immunol* 2011;32:345–9.
- 12 Zhu C, Anderson AC, Schubart A, et al. The Tim-3 ligand Galectin-9 negatively regulates T helper type 1 immunity. *Nat Immunol* 2005;6:1245–52.
- 13 Zhang R, Chen S, Luo T, et al. Activated Tim-3/Galectin-9 participated in the development of multiple myeloma by negatively regulating Cd4 T cells. *Hematology* 2024;29:2288481.
- 14 Ni X, Wu W, Sun X, et al. Interrogating glioma-M2 macrophage interactions identifies Gal-9/Tim-3 as a viable target against PTEN-null glioblastoma. *Sci Adv* 2022;8:eab15165.
- 15 Pang N, Alimu X, Chen R, et al. Activated Galectin-9/Tim3 promotes Treg and suppresses Th1 Effector function in chronic lymphocytic leukemia. *FASEB J* 2021;35:e21556.
- 16 Yang R, Sun L, Li C-F, et al. Galectin-9 interacts with PD-1 and TIM-3 to regulate T cell death and is a target for cancer Immunotherapy. *Nat Commun* 2021;12:832.
- 17 Limagne E, Richard C, Thibaudin M, et al. Tim-3/Galectin-9 pathway and mMDSC control primary and secondary resistances to PD-1 blockade in lung cancer patients. *Oncoimmunology* 2019;8:e1564505.
- 18 Kikushige Y, Miyamoto T, Yuda J, et al. A TIM-3/Gal-9 Autocrine stimulatory loop drives self-renewal of human myeloid leukemia stem cells and leukemic progression. *Cell Stem Cell* 2015;17:341–52.
- 19 Zhao L, Cheng S, Fan L, et al. TIM-3: an update on Immunotherapy. *International Immunopharmacology* 2021;99:107933.
- 20 Gonçalves Silva I, Yasinska IM, Sakhnevych SS, et al. The Tim-3-Galectin-9 Secretory pathway is involved in the immune escape of human acute myeloid leukemia cells. *EBioMedicine* 2017;22:44–57.
- 21 Rangachari M, Zhu C, Sakuishi K, et al. Bat3 promotes T cell responses and autoimmunity by repressing Tim-3-mediated cell death and exhaustion. *Nat Med* 2012;18:1394–400.
- 22 Oomizu S, Arikawa T, Niki T, et al. Galectin-9 suppresses Th17 cell development in an IL-2-dependent but Tim-3-independent manner. *Clin Immunol* 2012;143:51–8.
- 23 Madireddi S, Eun S-Y, Lee S-W, et al. Galectin-9 controls the therapeutic activity of 4-1Bb-targeting antibodies. *J Exp Med* 2014;211:1433–48.
- 24 Daley D, Mani VR, Mohan N, et al. Dectin 1 activation on macrophages by Galectin 9 promotes pancreatic carcinoma and peritumoral immune tolerance. *Nat Med* 2017;23:556–67.
- 25 Li Y, Feng J, Geng S, et al. The N- and C-terminal carbohydrate recognition domains of Galectin-9 contribute differently to its multiple functions in innate immunity and adaptive immunity. *Mol Immunol* 2011;48:670–7.
- 26 Stillman BN, Hsu DK, Pang M, et al. Galectin-3 and Galectin-1 bind distinct cell surface glycoprotein receptors to induce T cell death. *J Immunol* 2006;176:778–89.
- 27 Demotte N, Wieërs G, Van Der Smissen P, et al. A Galectin-3 ligand corrects the impaired function of human Cd4 and Cd8 tumor-infiltrating lymphocytes and favors tumor rejection in mice. *Cancer Res* 2010;70:7476–88.
- 28 Gordon-Alonso M, Hirsch T, Wildmann C, et al. Galectin-3 captures interferon-gamma in the tumor matrix reducing Chemokine gradient production and T-cell tumor infiltration. *Nat Commun* 2017;8:793.
- 29 Raiter A, Lipovetsky J, Stenbac A, et al. TNBC-derived Gal3Bp/Gal3 complex induces immunosuppression through Cd45 receptor. *Oncoimmunology* 2023;12:2246322.
- 30 Kouo T, Huang L, Pucsek AB, et al. Galectin-3 shapes antitumor immune responses by suppressing Cd8 T cells via LAG-3 and inhibiting expansion of Plasmacytoid Dendritic cells. *Cancer Immunol Res* 2015;3:412–23.
- 31 Bae J, Accardi F, Hideshima T, et al. Targeting Lag3/GAL-3 to overcome immunosuppression and enhance anti-tumor immune responses in multiple myeloma. *Leukemia* 2022;36:138–54.
- 32 Wu X, Giobbie-Hurder A, Connolly EM, et al. Anti-CTLA-4 based therapy elicits humoral immunity to Galectin-3 in patients with metastatic Melanoma. *Oncoimmunology* 2018;7:e1440930.
- 33 Lau KS, Partridge EA, Grigorian A, et al. Complex N-Glycan number and degree of branching cooperate to regulate cell proliferation and differentiation. *Cell* 2007;129:123–34.
- 34 Curti BD, Koguchi Y, Leidner RS, et al. Enhancing clinical and immunological effects of anti-PD-1 with Belapectin, a Galectin-3 inhibitor. *J Immunother Cancer* 2021;9:e002371.
- 35 Sun L, Li C-W, Chung EM, et al. Targeting Glycosylated PD-1 induces potent antitumor immunity. *Cancer Res* 2020;80:2298–310.
- 36 Pedersen K, Nielsen MA, Juul-Madsen K, et al. Galectin-3 interacts with PD-1 and Counteracts the PD-1 pathway-driven regulation of T cell and Osteoclast activity in rheumatoid arthritis. *Scand J Immunol* 2023;97:e13245.
- 37 Scafetta G, D'Alessandria C, Bartolazzi A. Galectin-3 and cancer Immunotherapy: a glycobiochemical rationale to overcome tumor immune escape. *J Exp Clin Cancer Res* 2024;43:41.
- 38 Rubinstein N, Alvarez M, Zwirner NW, et al. Targeted inhibition of Galectin-1 gene expression in tumor cells results in heightened T cell-mediated rejection: a potential mechanism of tumor-immune privilege. *Cancer Cell* 2004;5:241–51.
- 39 Toscano MA, Bianco GA, Ilairegui JM, et al. Differential Glycosylation of Th1, Th2 and TH-17 effector cells selectively regulates susceptibility to cell death. *Nat Immunol* 2007;8:825–34.
- 40 Ilairegui JM, Croci DO, Bianco GA, et al. Tolerogenic signals delivered by Dendritic cells to T cells through a Galectin-1-driven Immunoregulatory circuit involving interleukin 27 And interleukin 10. *Nat Immunol* 2009;10:981–91.
- 41 Dalotto-Moreno T, Croci DO, Cerliani JP, et al. Targeting Galectin-1 overcomes breast cancer-associated immunosuppression and prevents metastatic disease. *Cancer Res* 2013;73:1107–17.
- 42 Croci DO, Cerliani JP, Dalotto-Moreno T, et al. Glycosylation-dependent lectin-receptor interactions preserve angiogenesis in anti-VEGF refractory tumors. *Cell* 2014;156:744–58.
- 43 Cagnoni AJ, Giribaldi ML, Blidner AG, et al. Galectin-1 fosters an immunosuppressive microenvironment in colorectal cancer by Reprogramming Cd8+ regulatory T cells. *Proc Natl Acad Sci U S A* 2021;118:e2102950118.
- 44 Rudjord-Levann AM, Ye Z, Hafkenscheid L, et al. Galectin-1 induces a tumor-associated macrophage phenotype and upregulates Indoleamine 2,3-Dioxygenase-1. *iScience* 2023;26:106984.
- 45 Jouve N, Despoix N, Espeli M, et al. The involvement of Cd146 and its novel ligand Galectin-1 in apoptotic regulation of endothelial cells. *J Biol Chem* 2013;288:2571–9.
- 46 Nambiar DK, Aguilera T, Cao H, et al. Galectin-1-driven T cell exclusion in the tumor endothelium promotes immunotherapy resistance. *J Clin Invest* 2019;129:129025:5553–67.
- 47 Zheng Y, Zhang H, Xiao C, et al. Klf12 overcomes anti-PD-1 resistance by reducing Galectin-1 in cancer cells. *J Immunother Cancer* 2023;11:e007286.
- 48 Wu X, Li J, Connolly EM, et al. Combined anti-VEGF and anti-CTLA-4 therapy elicits humoral immunity to Galectin-1 which is associated with favorable clinical outcomes. *Cancer Immunol Res* 2017;5:446–54.
- 49 Bannoud N, Stupirski JC, Cagnoni AJ, et al. Circulating Galectin-1 Delineates response to bevacizumab in melanoma patients and reprograms endothelial cell biology. *Proc Natl Acad Sci U S A* 2023;120:e2214350120.
- 50 Yeboah M, Papageorgiou C, Jones DC, et al. Lirb3 (Ilt5) is a myeloid cell checkpoint that elicits profound immunomodulation. *JCI Insight* 2020;5:e141593.
- 51 Yang T, Qian Y, Liang X, et al. Lirb4, an immune checkpoint on myeloid cells. *Blood Sci* 2022;4:49–56.
- 52 Huang R, Liu X, Kim J, et al. Lirb3 supports immunosuppressive activity of myeloid cells and tumor development. *Cancer Immunol Res* 2024;12:350–62.
- 53 Wang Y, Sun Y, Deng S, et al. Discovery of Galectin-8 as an Lirb4 ligand driving M-Mdscs defines a class of antibodies to fight solid tumors. *Cell Rep Med* 2024;5:101374.